

Review Article

Integrin Inhibition in the Tumor Microenvironment – more complex than expected

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Abstract

Tumor cell migration and invasion are critical steps in the metastatic cascade and depend on the interaction between tumor cells, the extracellular matrix (ECM) and the endothelial cells. Integrins are key receptors that link cells and ECM, acting as mechanical sensors of the cell microenvironment. Particularly, Arg-Gly-Asp (RGD)-binding integrins such as $\alpha v\beta 3$ and $\alpha 5\beta 1$ integrins are of special interest in cancer progression. Integrins also interact with growth factor receptors resulting in an important cross talking between intracellular signaling pathways. Studies have provided evidence of distinct roles for $\alpha v\beta 3$ and $\alpha 5\beta 1$ integrins during migration. Therefore, $\alpha v\beta 3$ and $\alpha 5\beta 1$ integrins became an attractive target for pharmacological inhibition in cancer therapy and metastasis prevention. Cilengitide, the first integrin inhibitor based on the RGD motif, is currently under clinical trial in cancer patients with limited success. Monoclonal antibodies to integrins also presented modest results. Therefore, efforts to achieve a better understanding on the integrin roles in cancer progression are needed. In the last few years, new mechanisms that may help to explain the lack of success of integrin inhibitors were described and are commented here.

Keywords: integrin, cancer, disintegrins

Introduction

Cancer is one of the major concerns related to human health, and metastasis, when occurs, is the main cause of deaths in patients with cancer. Despite all efforts of academic, governmental and private institutions, there are very few options to prevent or treat metastasis. [1] One of the reasons for this lack of success relays on the current limited knowledge on the cellular programs driving the process of metastasis. Recently, new mechanisms that allow prolonged tumor cell survival after loss of attachment to the ECM have been reported, including autophagy and entosis [2]. In addition to shedding some light in the knowledge of tumor progression, these mechanisms provided new targets and options for metastasis treatment. Here we review some key aspects of cell attachment/detachment to ECM by integrins in the context of tumor microenvironment. We will also comment on the results obtained so far with integrin-targeted anticancer therapy.

Tumor Microenvironment and the Integrins

In the past ten years, the microenvironment where tumor cells develop has achieved special importance mostly due to its pivotal role in tumor progression. The tumor microenvironment (TME) provides signals and several kinds of support from distinct cell types and from the extracellular matrix (ECM) [3, 4]. Signals from the TME comprise soluble factors released by stromal cells such as fibroblasts, stem and immune cells, blood vessels, products from proteolysis of ECM components and cytokines [5]. On the other hand, tumor cells release proteases, microvesicles and growth factors that affect and modify the

surrounding cells and the ECM. The TME also has a key role in the resistance to therapy due to a continuous communication with tumor cells and modulation of their responses [6, 7]. Integrins are among the crucial cell surface receptors in supporting the cross talk between cells and ECM, and therefore are critically involved in tumor progression [8].

Integrins are transmembrane receptors that support the adhesion of cells to the ECM [8]. Loss of integrin adhesion usually induces cell death unless cells can find new adhesion sites. Integrin are formed by heterodimers containing one α and one β subunits with the ability to recognize ECM components such as collagen (Col), fibronectin (FN) and laminin (LM) with high affinity [9]. There are 18 α subunits and 8 β subunits that can be combined in several ways to form distinct receptors with different specificity to ECM components. For instance, $\alpha 5\beta 1$ integrin is the main receptor for FN whereas vitronectin (VN) binds preferentially to $\alpha v\beta 3$ integrin. Both $\alpha 5\beta 1$ and $\alpha v\beta 3$ integrins recognize the tripeptide RGD motif within ECM proteins; however other integrins also binds to the RGD motif such as $\alpha v\beta 1$, $\alpha v\beta 5$, $\alpha v\beta 6$, $\alpha v\beta 8$, $\alpha 8\beta 1$, and the platelet fibrinogen receptor, $\alpha IIb\beta 3$ integrin [10]. One of the most interesting feature of integrins is the fact that, despite binding the same ligand, each one has its own cell-dependent pattern of expression and plays distinct roles in cell adhesion and migration [11].

Integrins in cell adhesion and migration

Integrin clustering and activation upon ligand binding triggers intracellular signaling pathways including the activation of several kinases like the focal adhesion kinase (FAK), mitogen activated

protein kinase (MAPK) and extracellular signal-regulated kinase (ERK) resulting in changes in cell behavior [8, 12]. Cell adhesion and migration are among the main cell abilities that are under strict integrin control and are crucial steps during tumor progression. Moving cells may change their integrin profile in response to modifications on the ECM milieu. In a FN-rich microenvironment, cells will depend mostly on integrins such as $\alpha 5\beta 1$ and $\alpha v\beta 3$ for motility and, despite their ability in binding to the same ligand, these two integrins have distinct and specific roles in cell migration [13].

Adhesion to fibronectin by $\alpha v\beta 3$ integrin supports extensive actin cytoskeletal reorganization resulting in a single large lamellipod with static cell–matrix adhesions at the leading edge [14]. On the other hand, cell adhesion by $\alpha 5\beta 1$ generates thin protrusions containing highly dynamic cell–matrix adhesions in multiple directions [14]. Therefore, these authors concluded that $\beta 1$ integrins support random migration, whereas $\beta 3$ integrins are related to persistent migration. In agreement with these data, we have demonstrated that blockage of $\alpha v\beta 3$ integrin by a *DisBa-01*, a recombinant RGD-disintegrin from snake venom, resulted in loss of directionality and decrease of speed migration [15]. Despite having the RGD adhesive motif, which is recognized by both $\alpha v\beta 3$ and $\alpha 5\beta 1$ integrins, the dissociation constant of *DisBa-01* for the $\alpha v\beta 3$ integrin is 100 times higher than for $\alpha 5\beta 1$ integrin [15]. These results confirm the role of $\alpha v\beta 3$ integrin in defining directionality of the cell movement.

Rosa-Cusachs et al., 2009 also provided evidence of distinct roles for $\alpha 5\beta 1$ and $\alpha v\beta 3$ integrin during cell adhesion/migration processes, with the demonstration that $\alpha 5\beta 1$ integrin is responsible for supporting high adhesion forces while $\alpha v\beta 3$ integrin starts talin-dependent mechanotransduction. However, these effects depend on the substrate where cells attach. Fibroblasts exhibit persistence migration on FN-coated surfaces [16]. Ligand-specific activation of $\alpha v\beta 3$ e $\alpha 5\beta 1$ integrins also confirmed the distinct roles of each integrin in cell adhesion. Activation of $\alpha v\beta 3$ integrins led to stabilization of peripheral focal adhesions while fibrillary structures were observed when $\alpha 5\beta 1$ integrins are activated [17]. Moreover, αv -class of integrins such as $\alpha v\beta 3$ were demonstrated to outcompete $\alpha 5\beta 1$ integrins in FN binding; however, after engagement, αv -class integrins cooperate with $\alpha 5\beta 1$ integrins to form additional adhesion sites consequently strengthening cell adhesion to FN [18].

Although they may bind the same ligand, $\beta 1$ and $\beta 3$ integrins have distinct and cooperative roles in mechanotransduction. By means of traction force microscopy, Millieux and colleagues demonstrated that deleting $\beta 3$ subunit increases traction forces, whereas the deletion of $\beta 1$ subunit results in a strong decrease of contractile forces [19].

Integrins and Growth Factor Receptors

The cross talk between integrin and growth factor receptor (GFR) signaling pathways is well documented in the literature in both normal cells and tumor cells. Endothelial cells seem to be highly sensitive to integrin activation of GFRs including the vascular endothelial growth factor receptor (VEGFR), epidermal growth factor receptor (EGFR), and the platelet-derived growth factor receptor (PDGFR). This cooperative process is crucial in physiological and tumor angiogenesis

and it is considered one key factor in tumor progression. The $\alpha 5\beta 1$ integrin was demonstrated to induce tumor cell proliferation by two pro-survival mechanisms including direct activation the EGFR signaling cascade and by signaling via continuous integrin-dependent activation of protein kinase B (PKB, also known as AKT) [20]. Moreover, EGFR overexpression led to inactivation of $\alpha 5\beta 1$ integrin in A431 squamous carcinoma cells, which would be interesting in order to prevent cell interaction with the ECM and therefore to avoid tumor cell proliferation. However, treatment of cells with EGFR kinase inhibitor resulted in reactivation of the integrin [21]. Since GFRs are usually overexpressed in many types of cancer cells, pro-survival signaling pathways from both GFR and integrins would be more efficiently impaired upon association of anti-integrin/anti-GFR treatments than either treatment alone [22].

VEGF binding to its receptors and co-receptors induce receptor homodimerization and heterodimerization, followed by activation of tyrosine kinases and signaling cascades, with the association of a set of adaptor proteins. The activation of these signaling pathways results in activation, migration and proliferation of endothelial cells, crucial steps for neoangiogenesis. In addition, mechanical forces such as shear stress may activate VEGFR2 similarly to ligand binding, leading to the activation of intracellular signaling pathways [23]. Integrins are proposed to act as co-receptors upon VEGF binding to VEGFRs, similarly to neuropilins and heparin sulfate proteoglycans in endothelial cells [24]. However, integrins seem to participate actively in the control of VEGFR signaling. Blocking $\alpha v\beta 3$ integrin by a RGD-antagonist downregulated the expression of VEGF, VEGFR1 and VEGFR-2 in endothelial cells but not in MDA-MB-231 breast tumor cells [25]. Contrastingly, we observed an increase in VEGF protein levels by human fibroblasts after treatment with the $\alpha v\beta 3$ integrin antagonist. In addition, we have also demonstrated that this RGD-antagonist inhibits endothelial in vitro cell migration and in vivo angiogenesis in mice [26, 27]. These results suggest that the $\alpha v\beta 3$ integrin is not only a co-receptor; instead, it is an active partner in the control of VEGF signaling in endothelial cells.

A key role for $\alpha 1\beta 1$ and $\alpha 2\beta 1$ integrins in the process of angiogenesis triggered by vascular endothelial growth factor (VEGF) was previously reported. Antibody antagonism of either integrin resulted in potent inhibition of VEGF-driven angiogenesis in mouse skin, reduced tumor growth and angiogenesis of human squamous cell carcinoma xenografts. [28]

Integrins and MMPs

Matrix metalloproteases (MMP) comprise a class of zinc-dependent enzymes responsible for ECM remodeling and degradation. MMPs are also involved in the processing of growth factors, cytokines and surface transmembrane proteins. MMPs are produced as inactive zymogens that can be activated by proteolytic removal of the pro-peptide domain by furin, or by autoproteolysis. To date, more than 24 MMPs are known, including secreted or membrane anchored forms, (membrane-type MMPs, MT-MMPs) that play crucial roles in the process of MMP activation. For instance, activation of pro-MMP-2 at the cell surface involves the formation of a trimolecular complex with membrane type-1 metalloprotease (MT-1-MMP) and tissue

inhibitor of metalloproteases-2 (TIMP-2). MMP-2 and MMP-9 are also known as gelatinases A and B, respectively, due to the ability to digest degraded forms of collagen. Gelatinases are characterized by the presence of a fibronectin-like domain that drives the enzyme to its ECM substrates. MMP-2 and MMP-9 are of particular interest in cell migration and consequently, in metastasis development. MMP-2 is constitutively produced by most cells; however, MMP-9 is expressed by only a few types of cells including neutrophils, macrophages and tumor cells, being induced in several pathological conditions such as tumor invasion [29, 30].

MMPs play key roles in tumor progression such as degrading ECM to allow migration of tumor cells to distant secondary sites, allowing endothelial cell migration and proliferation to produce tumor angiogenesis, and releasing growth factors from the ECM to provide constant survival and proliferation signals.

The role of integrins in controlling MMP activity is less studied. Previous studies demonstrated that interaction of fibronectin with $\alpha 4 \beta 1$ integrin upregulates MMP-9 expression by infiltrating leukocytes in damaged livers and the blockade of this interaction disrupted leukocyte migration [30]. MMP-2 activation upregulates VEGF-A expression in melanoma cells via an $\alpha \nu \beta 5$ integrin/phosphoinositide-3-kinase-dependent pathway [31]. The $\alpha \nu \beta 3$ integrin has been closely related to tumor progression and reduced patient survival rates in melanoma, colon carcinoma and breast cancer, increasing migration and invasion of tumor cells [32, 33]. Blocking $\alpha \nu \beta 3$ integrin inhibited MT-1-MMP-dependent activation of MMP-2 induced by collagen I; however, cells expressing high levels of $\beta 3$ integrin subunit have increased abilities of adhesion and migration [34]. Blocking $\alpha \nu \beta 3$ integrin in endothelial cells by a RGD-based antagonist completely abolished MMP-2 activity; in contrast, the same treatment increased almost twice the MMP-9 levels in the conditioned media from MDA-MB-231 breast tumor cells [25]. These results demonstrated that one single specific integrin inhibitor might induce different cell-dependent effects.

Integrins and tumor progression

The correlation of expression levels of $\alpha \nu \beta 3$, $\alpha \nu \beta 5$, $\alpha 5 \beta 1$, $\alpha 6 \beta 4$, $\alpha 4 \beta 1$, $\alpha \nu \beta 6$ and $\alpha \nu \beta 8$ integrins with metastasis and poor patient prognosis is well documented (reviewed by Nieberler et al., 2017). [35] One of the most studied integrin in tumor progression is the $\alpha 5 \beta 1$ integrin. Higher levels of $\alpha 5$ subunit were found than in normal tissue in patients with advanced renal cell carcinoma and correlated with tumor grade, metastasis development and reduced patient survival [36]. Expression of $\alpha \nu \beta 6$ is significantly associated with the progression of breast ductal carcinoma to an invasive form by a mechanism involving upregulation of MMP-9 and transforming growth factor- β (TGF- β) [37].

Previous reports associated $\beta 1$ and $\beta 3$ integrins with TGF- β stimulation of epithelial–mesenchymal transition (EMT) and breast tumor metastasis by means of a compensatory mechanism [38]. Inactivation of $\beta 1$ integrin impairs the TGF- β effect in promoting tumor cell migration; however, a strong compensatory upregulation of $\beta 3$ integrin restores the induction of the EMT phenotypes by TGF- β ,

indicating that the two integrins must be targeted to prevent tumor progression [38].

Integrin-targeted therapy

Cilengitide was one of the first antiangiogenic drugs directed to blockade of cell adhesion to the ECM by antagonizing $\alpha \nu \beta 3$ and $\alpha 5 \beta 1$ integrins (IC₅₀ for inhibition of cell adhesion of 0.2 and 11 nM, respectively). In phase I studies, patients with advanced solid tumors were treated with cilengitide without consistent results [39]. In another study, cilengitide was tested in association with cediranib, an inhibitor of VEGFR-associated tyrosine kinase. The association of cilengitide with the two drugs was well tolerated, however there were no changes in the survival rates [40, 41]. Cilengitide combined with temozolomide, an oral alkylating agent, did not increase the survival rates of glioblastoma patients [42]. Among antiangiogenic drugs, only the anti-VEGF-A bevacizumab increased disease-free survival time in patients with glioblastoma [43]. Recently, 12 patients with solid tumors such as breast cancer were treated with cilengitide, with partial positive response to the treatment and 05 had stable disease as the best response [44]. In summary, after 10 years of clinical assays with cilengitide for different tumors, results are still disappointing. Reasons for the lack of success may be related to the determination of the effective doses and time of administration. [45- 47] Other reason for lack of success may be the dose-dependent opposing effects of cilengitide related to tumor angiogenesis, with low doses being pro-angiogenic in contrast with anti-angiogenic effect of higher doses [48]. However, a better comprehension on the distinct roles of integrins in the context of complexity of the tumor microenvironment is needed before discarding integrin-targeted therapy.

Volociximab, a monoclonal antibody anti- $\alpha 5 \beta 1$ integrin, has been tested in at least 10 phase I and II clinical trials to treat some types of tumors, including advanced non-small cell lung cancer (NSCLC) and metastatic melanoma, among others. Volociximab was used either as single therapy or in combination with classical drugs such as carboplatin and paclitaxel. Preliminary results showed modest but relevant results such as an increase in median progression-free survival of 6.3 months and decreased levels of potential biomarkers of angiogenesis or metastasis after six cycles of treatment [49].

Etaracizumab, an anti- $\alpha \nu \beta 3$ integrin monoclonal antibody, decreased SKOV3ip1 ovarian tumor cell proliferation and invasion in vitro and resulted in about 50% of tumor weight decrease in mice [50]. Combination therapy with paclitaxel gave better results in decreasing tumor weight, and tumors showed reduced levels of p-Akt and p-mTOR; however, microvessel density of resected tumors after therapy were not decreased [50]. The $\beta 1$ integrin subunit has been associated to therapeutic resistance to trastuzumab (anti-EGFR/HER2 monoclonal antibody) and to lapatinib (an EGFR/HER2 tyrosine kinase inhibitor) of human epidermal growth factor receptor (HER)-2-positive breast tumor cells [51].

One of the most intriguing question in integrin-based anti-cancer therapy is the fact that the strong and positive results on inhibition of tumor progression in pre-clinical assays did not translate to clinical

assays. Integrin inhibitors including monoclonal antibodies and synthetic molecules showed disappointed results in patient survival time, disease stabilization and the development of metastasis [42, 48, 52]. One reason for the negative results may rely on the complexity of the mechanism of action of integrins, their ability to compensate each other and inducing an even worse phenotype. Deleting $\beta 1$ integrin was compensated by $\beta 3$ integrin, which stimulated metastasis in murine model of breast cancer [38].

Besides the compensatory mechanism, integrin inhibition should induce cell death by anoikis, a kind of apoptotic cell death that occurs due to the loss of cell attachment to the ECM [53]. However, ECM detachment results in antiapoptotic signals, as a defense mechanism against anoikis until cells be able to find a new place to attach again. Meantime, cells undergo autophagy, a cell process mostly triggered by the loss of integrin-mediated adhesion that allow cell survival during some time. However, prolonged detachment will later induce apoptosis by anoikis [54]. One of the most critical finding is that tumor cells usually develop resistance to anoikis due to a sustained autophagic response [54].

Entosis, an even more sophisticated cell survival mechanism, was described [55]. Entosis, also referred to as cell-in cell structures, or cell cannibalism, is triggered by loss of attachment to ECM, similarly to the process of autophagy. Entotic cells have been observed in many types of tumors and may be one of the reasons for the abnormal number of chromosomes found in most tumors [2]. Engulfed cells are alive and may divide inside the host, and in case of tumor cells, such process may occur indefinitely. Such mechanisms of tumor cell survival that happen upon ECM detachment may certainly contribute for the lack of success of integrin inhibitors in clinical trials.

More recently, the mechanism of vessel co-option was described as a mediator of resistance to anti-angiogenic therapy of breast tumor liver and lung metastasis [56]. Vessel co-option is an alternative pathway of tumor cells for obtaining nutrients from blood using the pre-existing vasculature without producing new vessels. Blocking VEGF/VEGFR signaling induces co-option and tumor growth in glioblastoma patients. [57] Since there is a close reciprocal stimulatory relationship between VEGF and $\alpha v\beta 3$ integrin [58], one might expect a role for integrin inhibition in supporting the mechanism of co-option. This possibility remains to be elucidated. These results demonstrate the complexity of the TME and its relevance in tumor progression.

New insights on integrin targeting

In spite of being extensively described, integrin inhibition in tumor microenvironment is still challenging and attractive. The failure of targeted therapies so far leads to deeper investigations about signaling pathways, endocytic trafficking and recycling of integrins [59, 60].

As a result, the attention that before was outside cell, turned to intracellular integrin fate affecting the overall cell behavior. It is known that integrin trafficking dictates the nature of Rho GTPase signaling during cytokinesis and cell migration [61]. FN binding promotes both accelerated internalization and ubiquitination of $\alpha 5\beta 1$ receptors. Subsequently, alterations in pH inside endosomal compartments will define either recycling or degradation of integrins

[59, 62] When $\alpha 5$ integrin suffers ubiquitination upon FN binding, the ESCRT machinery acts sorting this receptor to either multivesicular bodies, recycling or lysosomal degradation [59]. It was previously demonstrated that mutation on the ubiquitination site in cytoplasmic tail of $\alpha 5$ integrin causes a different sorting of fibronectin on cell, affecting fibroblast migration [63].

The multivesicular bodies are cell compartments containing intraluminal vesicles named exosomes that are secreted to the extracellular space by shedding from the plasma membrane, improving cell communication with the TME [64]. The presence of both integrin and FN inside multivesicular bodies had light to the hypothesis that these receptors might be present in vesicles in the extracellular environment. Sung et al demonstrated that in fact $\alpha 5$ integrin is secreted as an exosome cargo, and more than that, FN was also secreted in these vesicles. Thus, a new and promising science of integrins as a target has emerged from the microvesicles field. Studies on cell communication had proved that integrin transfer can occur through exosomes delivery in both TME and the pre-metastatic niche [65]. In addition, the integrin content of exosomes can change the types of integrin expression in the new tumor focus [66]. The real contribution of integrins on cell communication using cell-derived vesicles is still unclear, as well as the effects of blockage of these receptors. However, this can be the missing clue that can explain the failure of earlier integrin inhibitors in drug development.

Conclusions

Cell attachment to the ECM via integrins is a key factor for tumor progress; however, integrin inhibition may be carefully considered as a target for drug development. Combination of anti-growth factors antibodies, tyrosine kinase inhibitors and integrin inhibitors may be an interesting choice but must be first evaluated in pre-clinical assays considering all the possible escape mechanisms that tumor cells can develop. Most of the studies so far have considered the signaling in a cellular level, however, in complex organisms, several other factors might systemically interfere on integrin inhibition, making the drug development even more challenging. The integrins may have won some battles but not the war.

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